

Making Tick Antiserum

..... Safer for Cats



Purified Tick Antiserum for Cats

KEY FACTS

- Highly purified & concentrated paralysis tick antiserum
- Use to reduce tick antiserum reaction risk in cats
- Convenient individually boxed SINGLE DOSE vials
- APVMA Permit approved for use in cats
- New product in 2022

For more information contact:
info@padulaserums.com.au

Reference: Padula AM. (2022) Safety and efficacy of a purified canine immunoglobulin G formulation for treatment of 76 cats clinically affected by the Australian paralysis tick (*Ixodes holocyclus*). Aust Vet J (accepted in press)

APVMA Permit No. 90764

CAUTION

KEEP OUT OF REACH OF CHILDREN
READ SAFETY DIRECTIONS
FOR ANIMAL TREATMENT ONLY



PURIFIED TICK ANTISERUM FOR CATS

FOR ANIMAL TREATMENT ONLY

This product is NOT registered and is only available under APVMA permit number 90764 to a registered veterinarian.

CONSTITUENT STATEMENTS

Anti-Ixodes holocyclus immunoglobulin (>500 units/mL). Contains no preservative.

CLAIMS

This highly purified tick antiserum helps to reduce the clinical signs of tick paralysis in cats caused by Ixodes holocyclus by neutralising circulating toxin.

NET CONTENTS

5 mL

DIRECTIONS FOR USE

READ THE ENCLOSED LEAFLET BEFORE USING THIS PRODUCT

To be used by, or under the direct supervision of, a registered veterinary surgeon.

PRECAUTIONS

PRECAUTIONS IN ADMINISTRATION OF PURIFIED TICK ANTISERUM

Purified Tick Antiserum is a highly purified immunoglobulin product derived from dogs and is foreign protein when administered to cats. There is still potential for an acute anaphylactoid response to intravenous administration however this risk is low due to the purification process.

It is recommended to pre-medicate the patient

with dexamethasone and antihistamine to minimise the risk of adverse reactions.

Purified Tick Antiserum should be diluted in sterile saline diluent, warmed to body temperature, and administered by slow intravenous infusion over 20 minutes. Administration should cease if adverse reaction occurs, in which case consider administration of adrenaline, but re-administration by slow intravenous infusion may be possible.

SIDE EFFECTS

Facial swelling (occasional). Anaphylaxis (rare).

DOSAGE AND ADMINISTRATION

Purified Tick Antiserum should be administered as soon as possible after envenomation has occurred.

DIAGNOSIS OF TICK PARALYSIS

Diagnosis is usually based on clinical signs of lower motor neurone paralysis, characteristic respiratory distress, and confirmed where possible by the presence and species identification of an engorged paralysis tick.

SUPPORTIVE CARE FOR CATS WITH TICK PARALYSIS

Purified Tick Antiserum should not be considered the only treatment. Supportive care of affected cats is important for uncomplicated recovery. In severely affected cases mechanical ventilation may be required to support breathing or other forms of oxygen supplementation for those less severely affected. Cats should be hospitalised, stress minimised, and supported with appropriate fluid and nutritional therapy. Antibiotics may be indicated to minimise infection from aspiration events. Cats should be repeatedly searched for ticks as missed ticks can be fatal. Appropriate acaricidal treatment should also be administered at the discretion of the attending veterinarian.



INITIAL DOSING

All cases should initially receive no less than one vial of Purified Tick Antiserum. Large cats (>6 kg) and severely affected cats should preferably receive two vials until further data on dosing studies is available.

The time from administration of tick antiserum to clinical improvement has been reported as a minimum of 12 hours. Cats that have previously had tick antiserum or canine blood products are at higher risk of anaphylaxis to repeat administration. Treatment with a single large dose of Purified Tick Antiserum as early as possible in the disease process is recommended rather than attempting to titrate the dose to clinical effects.

GENERAL DIRECTIONS

Purified Tick Antiserum is prepared by extensive purification of canine tick antiserum. The canine tick antiserum is fractionated, and a highly purified immunoglobulin concentrate prepared containing negligible canine serum albumin and other non-immunoglobulin proteins.

FURTHER INFORMATION

Leister, E., J. Morton, R. Atwell and R. Webster (2018). Clinical presentations, treatments and risk factors for mortality in cats with tick paralysis caused by *Ixodes holocyclus*: 2077 cases (2008-2016). *J Feline Med Surg* 20(6): 465-478.

Padula, A. M. (2022). Safety and efficacy of a purified canine immunoglobulin G formulation for treatment of 76 cats clinically affected by the Australian paralysis tick (*Ixodes holocyclus*). *Aust Vet J* (accepted June 2022)

Padula, A. M., E. M. Leister and R. A. Webster (2020). Tick paralysis in dogs and cats in Australia: treatment and prevention deliverables from 100 years of research. *Aust Vet J* 98(1-2): 53-59.

SAFETY DIRECTIONS

Care should be taken to avoid accidental self-injection.

FIRST AID INSTRUCTIONS

In the event of accidental self-injection, seek medical advice immediately.

ADDITIONAL USER SAFETY

While this product is well tolerated by cats, there is a risk of serious adverse effects in humans associated with accidental self-injection.

DISPOSAL

Dispose of empty containers, outer packaging or expired product by wrapping with paper and putting in garbage.

Discarded needles should immediately be placed in a designated and appropriately labelled 'sharps' container.

STORAGE

Store between 2°C and 8°C (Refrigerate. Do not freeze.) Protect from light.

NAME & ADDRESS

Padula Serums Pty Ltd
100 Bosworth Road
Bairnsdale VIC 3875
AUSTRALIA

APVMA Lic. No. 1123



Purified Tick Antiserum Infusion Record

Clinic Name		Vet	
Date		Patient Name	
PTAS Batch	PTAS-7	No. Vials	
Prior Hx of TAS	Yes No Unknown	Total Diluted Volume	

Pre-Medication

Drug(s)	Dose	Route	Time Given

Clinical Assessment

Gait Score				Respiratory Score				No. Ticks Removed	Tick Clip Performed	Acaricide Treatment
1	2	3	4	A	B	C	D			

Gait Score: (1) Mild weakness; (2) Can stand but not walk; (3) Cannot stand but can right itself and maintain sternal recumbency; (4) Unable to right itself, cannot maintain sternal recumbency.

Respiratory Score: (A) Normal; (B) Mild: increased respiratory rate & effort; (C) Moderate: any respiratory distress or dyspnoea, restrictive breathing pattern, coughing, gagging or retching; (D) Severe: severe dyspnoea, cyanosis, progressive reduction in respiratory rate, open mouth breathing.

Infusion Observations

Time	Volume infused	Infusion rate	HR	RR	Resp Effort	MM Colour	Comments

Comments

TO HELP MAKE THIS A BETTER PRODUCT PLEASE SCAN INUFSION RECORD + CASE HISTORY AND SEND TO: Dr Andrew Padula info@padulaserums.com.au





SHORT COMMUNICATION

Safety and efficacy of a purified canine immunoglobulin G formulation for treatment of 76 cats clinically affected by the Australian paralysis tick (*Ixodes holocyclus*)

AM Padula^{a,b,*}

Acute adverse reactions in cats administered unrefined canine paralysis tick (*Ixodes holocyclus*) antiserum are commonly observed by veterinarians and can lead to significant morbidity and potentially fatal. A purified antiserum canine IgG concentrate was chromatographically prepared and aseptically formulated in single doses containing the equivalent of 5 mL of unrefined tick antiserum (TAS). The IgG was used for slow intravenous infusion into clinically affected cats at multiple veterinary clinics on the eastern seaboard of Australia. Overall, 72/76 (95%) of cats survived hospital discharge, an efficacy comparable to published data. A subset of 22 cats previously treated with unrefined TAS and considered high risk were included in the dataset. The safety profile was excellent with 0/76 acute adverse reactions although 2/76 (2.6%) developed mild facial swelling within 2 h of infusion that responded to the antihistamine. In conclusion, cats intravenously infused with purified IgG from canine TAS did not exhibit the expected frequency of acute adverse reactions during infusion and it was both safe and effective for the treatment of tick paralysis in cats.

Keywords cats; immunoglobulin; *Ixodes holocyclus*; tick antiserum; tick paralysis

Abbreviations IgG, immunoglobulin G; PTAS, purified tick antiserum; TAS, tick antiserum

Aust Vet J 2022

doi: 10.1111/avj.13194

Cats frequently succumb to lower motor neurone paralysis from salivary gland neurotoxins found within engorged Australian paralysis ticks (*Ixodes holocyclus*).¹ Alongside supportive hospital care, the core treatment for clinically affected cats is the administration of canine origin tick antiserum (TAS) to neutralise circulating tick neurotoxins.² Despite the apparent benefit of TAS, acute and sometimes fatal adverse reactions occur within minutes of administration and were reported to occur in 161/1735 (9.3%)³ and 375/6074 (6.2%)⁴ of cats.

The clinical description of acute TAS reactions in cats appears primarily anaphylactic in nature.⁵ The signs described include rapid onset pawing at the face, gagging, vomiting, salivation, piloerection on the back of the neck, tachycardia, collapse, severe respiratory distress and cardiac arrest. Clinical signs in cats sensitised to bovine serum albumin after intravenous challenge were similar to that of an acute TAS reaction and had a fatal outcome.⁶

The potential for acute adverse reactions during TAS administration to cats is recognised amongst veterinary practitioners and occurs despite premedication with corticosteroids, antihistamines and adrenaline.^{7, 8} Cats that experienced a TAS reaction were reported to have a higher risk of mortality overall and risk versus benefit approach was advocated for its use.³ Previous treatment with TAS appears to increase the risk of an acute reaction occurring, likely through sensitisation to the foreign protein.³ TAS reactions in cats were associated with 5.3 times increased incidence of mortality by day 5 of hospitalisation in the largest study to date in cats.³

The problem of acute adverse reactions to antiserum administration is not unique to TAS in cats and a similar problem was noted over 100 years ago during the initial development of serum therapy against diphtheria toxin in humans.⁹ Unrefined horse serum from donor horses immunised against diphtheria toxin was widely used to treat children and although spectacularly successful in some cases, acute reactions and deaths within minutes of administration soon became recognised as a significant problem.⁹ However, a reduction in the frequency of acute reactions to heterologous antiserum administration occurred when crude serum was fractionated into purified immunoglobulin (IgG) formulations consisting of either purified whole IgG or enzymatically digested into F(ab')₂ fragments.¹⁰ Experimental studies in guinea pigs, a species highly sensitive to sensitisation and anaphylaxis, demonstrated that removal of non-IgG protein by a two-step purification process almost entirely prevented anaphylaxis following intravenous challenge.¹¹ Commercially available TAS in Australia is unrefined crude serum and methods of production have changed little since the first published method of production.¹²

This study was undertaken to test the hypothesis that fractionation of TAS into a highly purified immunoglobulin formulation would reduce the risk of acute adverse reactions when administered intravenously to cats whilst retaining efficacy.

Commercial TAS was purchased and purified using a two-step process consisting of chemical precipitation¹³ of non-IgG protein

*Corresponding author.

^aPadula Serums Pty Ltd, Bairnsdale, Victoria, 3875, Australia; info@padulaserums.com.au^bAustralian Venom Research Unit, Department of Pharmacology and Therapeutics, Faculty of Medicine, Dentistry and Health Science, University of Melbourne, Parkville, Victoria, 3010, Australia

followed by chromatographic adsorption on Diethylaminoethyl Sephadex.¹¹ The finished product was extensively diafiltered against 0.01 M phosphate buffered saline pH 7.4 to remove phenol and other low molecular weight substances, concentrated by

ultrafiltration, sterile filtered and aseptically dispensed into sterile 10 mL glass vials presented as single doses. Potency testing was performed by indirect ELISA¹⁴ using a partially purified *I. holocyclus* neurotoxin adsorbed antigen. Endotoxin, pH, sterility, total IgG and purity of the final bulk were assessed by standard pharmacopeial methods. Preservative-free single-dose vials were prepared to contain not less than 2,500 units/vial of anti-*I. holocyclus* IgG in a volume of 5 mL. The total unit was chosen to be equivalent in neutralising capacity to 5 mL of TAS.

Veterinarians were invited to participate in the study of purified tick antiserum (PTAS) in cats, to directly substitute the PTAS-like-for-like with TAS and to follow their usual clinical treatment protocol. A dose recommendation of one vial was made and administration of additional vials was made at the discretion of the attending veterinarian. Clinical severity scores of gait and respiration were performed 1–4 by the clinician.³ A detailed record during each PTAS intravenous infusion of heart rate, respiratory rate, and SPO₂ was made. Acute adverse reactions were defined as those occurring within 15 min of infusion and delayed reactions within 2 h. Veterinarians submitted clinical notes shortly after treating each case, PTAS infusion records and feedback. The study was conducted under the remit of the Australian Pesticides and Veterinary Medicines Authority (APVMA) research permit PER7250.

The PTAS purification process was reproducible. The final IgG formulation migrated on an agarose gel as a single peak in the gamma globulin zone with the complete absence of canine serum albumin (Figure 1). The visual appearance of PTAS in the vial was a colourless, clear and particle-free liquid.

Survival rates at discharge varied as expected with severity (Table 1) but were unaffected by a dose of PTAS (Table 2). One cat died in

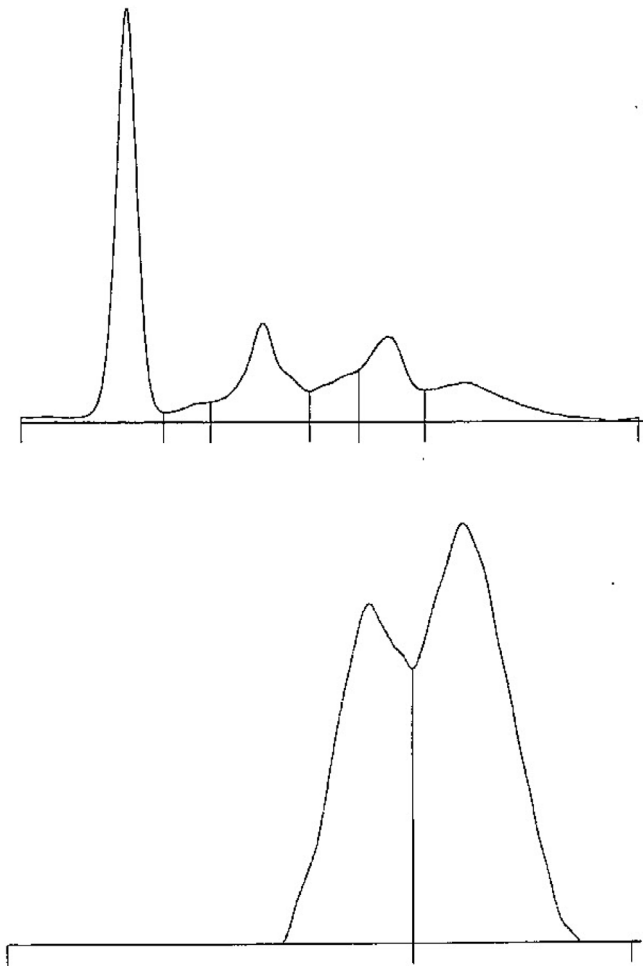


Figure 1. Agarose gel electrophoresis profiles demonstrated complete removal of albumin from TAS. (Top) serum protein migration of unrefined TAS and (bottom) PTAS with the migration of gamma globulin only. Albumin migrates strongly towards the positive electrode and is the first tall peak on the left side of unrefined TAS. PTAS, purified tick antiserum; TAS, tick antiserum.

Table 2. Dose of purified tick antiserum and case outcome

No. vials administered	No. cases (% total)	No. survived to discharge (%)
1	39 (51)	38 (97)
>1	36 (47)	33 (92)
NR	1 (1.3)	1 (100)
Total	76 (100)	72 (95)

Table 1. Survival to hospital discharge, incidence of acute and delayed reactions in cats receiving purified tick antiserum

Gait score	No. cases (% total)	No. survived to discharge (%)	No. acute reaction (%)	No. delayed reaction (% total)
1	22 (29)	22 (100)	0 (0)	1 (1.3)
2	30 (39)	29 (97)	0 (0)	1 (1.3)
3	19 (25)	18 (95)	0 (0)	0 (0)
4	2 (2.6)	0 (0)	0 (0)	0 (0)
NR	3 (3.9)	3 (100)	0 (0)	0 (0)
Total	76 (100)	72 (95)	0 (0)	2 (2.6)

Percentages rounded up. NR, not recorded.

hospital and three cats were euthanised on cost and prognosis. No acute adverse reactions were observed during PTAS infusion although two cats did develop delayed facial swelling which responded to antihistamine treatment. A subset of 22 cats treated with PTAS had a prior history of previously receiving TAS on 1–3 occasions and were speculated to be at a higher risk for a reaction, however, no acute reactions were observed.

The safety profile was excellent with no acute adverse reactions observed in 76 cats treated with 1 to 2 vials. Veterinary practice protocols varied in the duration of infusion and ranged between 30 to 240 min. Facial swelling, presumably caused by angioedema, is a recognised issue with homologous and heterologous plasma transfusion in cats and dogs.¹⁵ An overall survival rate of 97%³ was reported for 2077 cats treated for tick paralysis which is similar to the 95% observed here for PTAS. Administration of TAS to cats was reported to increase survival rates although cats that experienced an acute adverse reaction were more likely to die.³ Cats readily make IgG to canine albumin (Padula, unpublished data) contained in TAS. It is likely that IgE specific to canine albumin is also generated and sensitises cats to subsequent anaphylaxis; although fatal acute reactions have been reported in cats that have never been treated with TAS.³

Cats previously treated with unrefined TAS should based on theory, be at higher risk of acute anaphylactic reactions to subsequent TAS infusions. Although the dataset presented here is small, cats previously treated with TAS experienced no acute adverse reactions to PTAS. This observation is encouraging and supports the use of PTAS in this potentially higher-risk group has to merit in reducing infusion risk. However, there may be benefits other than explicitly in cats that have had prior TAS treatment because prior exposure does not explain those adverse reactions reported in naïve cats. Further studies to examine specific IgE responses and their role in acute reactions are required.

A number of APVMA licensed highly effective tick paralysis preventatives have become available since 2018 to cat owners and veterinarians, focusing the clinical need on refinement of ever more effective and safer treatments for those feline cases that do present to veterinarians.²

This study has provided preliminary evidence that the use of PTAS in cats can prevent acute adverse reactions from occurring compared to rates in published data, without compromising efficacy.

Acknowledgments

The author acknowledges the assistance and enthusiasm of the following veterinary clinics for participating in this study: Animal Emergency Services, Underwood, Qld; PetICU, Underwood, Qld;

Northside Emergency Veterinary Service, Sydney, NSW; Emergency Vets 24/7, Cairns, Qld; Merimbula & Pambula Veterinary Clinic, Merimbula, NSW and Sydney Animal Hospitals, Avalon, NSW. Open access publishing facilitated by The University of Melbourne, as part of the Wiley - The University of Melbourne agreement via the Council of Australian University Librarians.

Conflicts of interest and sources of funding

PTAS was manufactured by Padula Serums, a company owned by the author Andrew Padula. Boehringer Australia generously provided funding to undertake this research with the endorsement of the Australian Paralysis Tick Advisory Panel.

References

1. Padula AM. Tick paralysis of animals in Australia. In: Gopalakrishnakone P, Faiz SMA, Gnanathasan CA, Habib AG, Fernando R, Yang CC, editors. *Clinical toxicology*. Springer, Netherlands, Dordrecht, 2016;1–20.
2. Padula AM, Leister EM, Webster RA. Tick paralysis in dogs and cats in Australia: treatment and prevention deliverables from 100 years of research. *Aust Vet J* 2020;98:53–59.
3. Leister E, Morton J, Atwell R et al. Clinical presentations, treatments and risk factors for mortality in cats with tick paralysis caused by *Ixodes holocyclus*: 2077 cases (2008–2016). *J Feline Med Surg* 2018;20:465–478.
4. Atwell RB, Campbell FE. Reactions to tick antitoxin serum and the role of atropine in treatment of dogs and cats with tick paralysis caused by *Ixodes holocyclus*: a pilot survey. *Aust Vet J* 2001;79:394–397.
5. Schull D. Acute side effects attributed to the use of tick antitoxin serum: a review of available descriptions. *Aust Vet Practitioner* 2007;37:98.
6. McCusker H, Aitken I. Anaphylaxis in the cat. *J Pathol Bacteriol* 1966;91:282–285.
7. Fitzgerald M. *Ixodes holocyclus* poisoning. Sydney Postgraduate Foundation Course Proceedings, *Clinical Toxicology* 1998:203–220.
8. Malik R. Tick paralysis in the cat. Sydney Postgraduate Foundation Course Proceedings, *Clinical Toxicology* 1998:147–148.
9. Weaver G. Serum disease. *Arch Intern Med* 1909;3:485.
10. Gronski P, Seiler FR, Schwick HG. Discovery of antitoxins and development of antibody preparations for clinical uses from 1890 to 1990. *Mol Immunol* 1991;28:1321–1332.
11. Szegli G, Toma E, Gartner M. Removal of non-antibody protein from purified antitoxin sera by adsorption with DEAE Sephadex. *Rev Roum Biochim* 1968;5:157–160.
12. Oxer D. The preparation of canine anti-tick serum. *Aust Vet J* 1948;24:95–96.
13. Rojas G, Jimenez JM, Gutierrez JM. Caprylic acid fractionation of hyper-immune horse plasma: description of a simple procedure for antivenom production. *Toxicon* 1994;32:351–363.
14. Hall-Mendelin S, O'Donoghue P, Atwell RB et al. An ELISA to detect serum antibodies to the salivary gland toxin of *Ixodes holocyclus* Neumann in dogs and rodents. *J Parasitol Res* 2011;2011:283416.
15. Blois SL. Transfusion-associated complications. In: Yagi K, Holowaychuk M, editors. *Manual of veterinary transfusion and blood banking*. First edn. USA: John Wiley & Sons, 2016;155–171.

(Accepted for publication 11 June 2022)